Ozark Lichens — Basic Lichen IDs

For the Northern Arkansas Master Naturalists Program

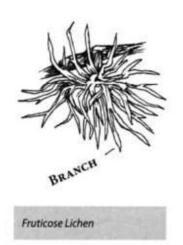
By Nastassja Noell

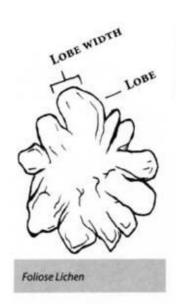
Key and species description excerpts by Douglas Ladd from "Ozark Lichens: Enumerating the lichens of the Ozark Highlands of Arkansas, Illinois, Kansas, Missouri, and Oklahoma" April 2018 Draft

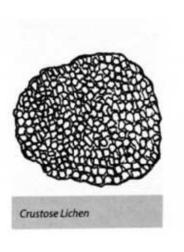
Textbook excerpts from "Radical Lichenology" by N. Noell in *Radical Mycology* by Peter McCoy (Chthaeus Press 2016)

Basic Morphological Characters

Basic Morphotypes



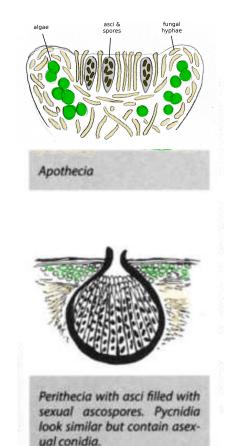




Asexual / Diaspore type:



Sexual / Fruiting Body:

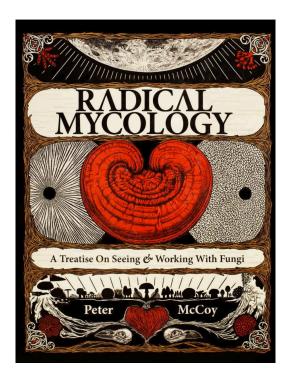


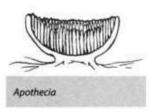
Key

Algae or Cyanobacteria

Medulla (loose hyphae)

Cortex (dense hyphae)







Perithecia with asci filled with sexual ascospores. Pycnidia look similar but contain asexual conidia.

Excerpts from "Radical Lichenology" by N. Noell in Radical Mycology by Peter McCoy (Chthaeus Press 2016)

The Reproductive Structures of Lichens

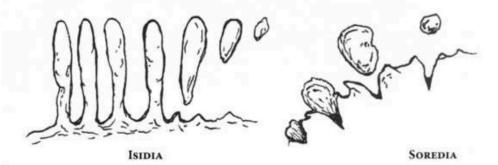
Though the algae and fungi in a lichen cohabitate, they do not share DNA. Both organisms reproduce independent of the other and a lichen as a whole may have multiple ways of replicating itself. The mycobiont tends to reproduce much like other fungi—often through the production of sexual spores and/or asexual conidia. The Ascomycete lichenized fungi have sexual reproduction patterns that tend to reflect those of their mushroom-forming kin, especially the Cup Fungi in the order Pezizales. In Ascomycete dominant lichens, the spore producing ascomata are generally apothecia or perithecia. Some Basidiomycete lichens do exist, however these are rare and often mistaken for mushrooms. The fruiting body structure in a Basidiomycete lichen is often similar to an agaric mushroom, however the mycelium and associated green algae form a distinct, superficial vegetative thallus (e.g. the basal scales of *Lichenomphalia hudsoniana*). The exception is the basidiolichen *Dictyonema s.l.* which looks and feels like a polypore mushroom, but the photobiont lives in the interior of the thallus.

After sporulation by the ascomata or basidiomata, the spores grow independently for a short while until an appropriate photobiont is found. These spores take a variety of shapes, sizes, colors, and form. As with mushrooms, the general spore types are usually consistent across genus or family. Most lichens also have asexual fruiting bodies (conidiomata) that produce conidia. In some species, conidia have been found to act as spermatia, fertilizing another lichen through a structure known as trichogyne.

Interestingly, most lichens also feature one or several methods for asexual cloning of the lichen itself. Instead of, for example, producing new spores with unique DNA sequences, these lichen clones are little bundles of fungal hyphae containing several photobiont cells. These bundles, called *diaspores*, bud off and fall away or are carried by the wind to new habitats where they serve as "seeds"—cottony clones of the mother lichen—that will eventually grow to be a new thallus. Diaspores can be dispersed hundreds of miles on the feet of traveling birds and in the air currents of the upper atmosphere, or more locally on the backs of insects and animals. Often the diaspores simply fall from the mother lichen to establish on a lower branch or below a host boulder.

When a diaspore lands on a suitable substrate and the right moisture and nutrients are present, it will first grow rhizomorph-like structures over the surface of the substrate. From this structure the thallus' tissues will begin to grow from the center outwards on top of the substrate, forming the cortices, the medulla, and a layer of photobiont cells. Diaspores come in two main forms:

- ISIDIA: These diaspores grow from within the medulla and push up through the
 cortex, bending the cortex around the diaspores, forming a protective cortex that
 then breaks off in finger-like pieces.
- SOREDIA: These diaspores lack any protective cortex. They are granular outgrowths
 of the medulla that grow up through openings in the cortex called soralia.



Diaspore-producing lichen species have distinct morphologies that aid in their identification. Lichens that produce isidia will not produce soredia, and vice versa. The diaspore type and location of origin are significant characteristics that likely reflect speciation events in the evolution of a particular group of lichens. Most sorediate and isidiate species will also occasionally still produce viable apothecia (in addition to their usual asexual diaspores), however there are a few species, e.g. *Lepraria spp.*, that have never been observed in the sexual state. Very rarely one will encounter sorediate or isidiate forms of species that normally do not produce diaspores. This terminology might be confusing at first, but just remember that diaspores contain both algal and fungal symbionts and thus reproduce the whole lichen as a clone, while spores reproduce only the mycobiont.

- for the forest. Nitrogen is a primary limiting nutrient in most ecosystems.
- · FOOD WEB: Lichens provide critical winter forage in temperate to boreal forests. Horse-Hair Lichens (Bryoria spp.) are the primary winter forage of keystone species including the woodland caribou. A variety of small mammals depend on them for food as well.
- · INSECTS: Insect-lichen associations are relatively unknown. Some insects such as Lacewings use lichens as camouflage, but there are likely many more intersections to be discovered.

Identifying Lichens

Before one can begin to work with lichens, it is essential to be able to first learn how to identify them. Identifying lichens is one of the most rewarding ways of engaging with lichens for it not only enhances personal and ecological resiliency but also increases one's connection to a habitat. As you learn how to identify lichens, more and more species begin to reveal themselves. A forest that previously looked like a wash of only one or two lichens soon turns into an ecosystem covered in hundreds of species.

Luckily, learning to identify most of the larger lichens is not too difficult and requires little equipment. If you're an herbalist, a good 10x or 20x loupe, field guide, and practice differentiating between look-alike species is all you will need. If you're an artist and want to collect dye lichens, you'll also need to do spot tests, as described below. If you're a citizen scientist doing environmental monitoring you'll probably also want a dissecting scope in order to identify lots of different species within a shorter period of time. And if you're a naturalist measuring total biodiversity, you'll eventually also want a compound microscope and the chemicals known as P and I.

Identifying lichens first begins with determining the overall structure of the lichen, generally classified by the following three forms:

- . FRUTICOSE: These lichens have a tree-like or beard-like form and are found in the greatest abundance growing in temperate rainforests. They tend to hang from trees where their large surface area is able to absorb as many nutrients and as much water from the air as possible. In more arid forests or areas with air pollution issues, fruticose lichens are often low in abundance and diversity. Unique features of fruticose lichens include branches and a uniform outer cortex (no distinction between upper and lower cortex is possible).
- FOLIOSE: Foliose lichens are flatter and more leaf-like. They come in a wide range of shapes and sizes and are often found in the greatest abundance in moist temperate forests on the bark and branches of trees and on top of moss at the bases of trees or rocks. Most are attached to the substrate by rhizines (short root-like structures) and the thallus usually forms a rosette (rose shapes), where each section is called a lobe. Lobes can be elongated like fingers or squat like rose petals. Lobes that are smaller than 2 millimeters in length are called squamules; lichens with many squamules are called squamulose. Unique features of foliose lichens include lobes, differentiated upper and lower cortices (usually both are present) and rhizines.
- · CRUSTOSE: These lichens are the most diverse group of lichens. They are found growing in all habitats, from the bark in tropical rainforests to the soil of arid deserts to frigid rocks in Antarctica. These lichens grow along or within the surface of their substrate, forming a living skin that facilitates water absorption and erosion prevention in desert habitats while also providing an anti-herbivory shield for thin barked trees in temperate and tropical forests. Unique features of crustose lichens include an upper cortex (no lower cortex) and areoles (the tile-like subunits making up the thallus of many crustose lichens).

Excerpts from "Radical Lichenology" by N. Noell in Radical Mycology by Peter McCov (Chthaeus Press 2016)

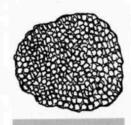


Fruticose Lichen



Foliose Lichen





Crustose Lichen

THE LICHEN RAINBOW

Are lichenologists colorblind? Sometimes it sure seems like it! Describing the colors of lichens for identification purposes is a highly subjective, and rather contentious topic among both amateurs and professionals. Is a lichen containing usnic acid called yellow, or yellow-green, or pale green? Ask three lichenologists and you might get three different answers. Similarly, a lichen containing the compound atranorin may be called blue by one person or white by another. It's all a bit ridiculous, but the matter is more confounded by the fact that lichen colors tend to vary when they are wet, dry, shaded, or exposed to the sun. Thus, some tips are offered to aid in determining a lichen's color:

- · Try to ID lichens only when they are dry. This is when their pigments are most visible and consistent.
- · Learn to recognize lichen pigments instead of colors. Begin associating the color you see with the chemical produced by the lichen, that way you can learn the range of color variation of "usnic green," "atranorin gray," etc.
- . When collecting lichens remember to note if the lichen was in a shaded location - lichens exposed to less sunlight produce less pigment and are thus more pale or almost green colored.

Spot Tests

As with identifying mushrooms, proper identification of a lichen may require the use of color change-inducing chemical reagents. This process is slightly different from that of working with mushrooms, but with some practice it can often be done quickly in the field. The materials for spot tests include:

- 2-4 SMALL GLASS CONTAINERS: These are for holding the chemicals. I prefer glass tincture bottles with eyedroppers that seal at the top.
- . CHEMICAL APPLICATION DEVICE: I prefer glass capillary tubes, others use a dissecting probe. Eyedroppers apply too much chemical, producing inaccurate reactions.
- · RAZOR BLADE
- DISSECTING MICROSCOPE OR LARGE MAGNIFYING GLASS
- · CHEMICALS: The most commonly used chemicals for lichen identification are K (10% potassium hydroxide KOH) and C (normal household bleach). As you get more comfortable with lichen identification you will want to add E (ethanol or methanol at 70% or higher), P (p-Phenylenediamine), and I (Lugols iodine) to your repertoire.
- UV LAMP: Centered on 350 nm (see below).

Spot tests often need to be applied to both the cortex and the medulla of the lichen, and often in a specific order, so make sure the capillary tubes are specific to only one chemical. I accomplish this by making my K tube longer than my C tube since KOH is more commonly called for in most ID keys. To limit having the toxic P test rolling around, I make the P tube so long that it rests in the P mixing container.

A UV lamp is also important for identifying lichens in tropical or subtropical areas, less so in temperate areas. Tropical lichens often contain xanthones, subtle yellowish pigments that fluoresce under UV light. UV lamps are also useful for other groups of lichens, including Cladonia and Parmotrema. The lamp must emit UV with a wavelength of around 350 nm in order for most UV+ substances to fluoresce. Cheap UV LED flashlights do not work! Some experimentation may be required to find a suitable lamp. 16 To conduct a UV spot test, simply go into a dark closet or cut holes out of a cardboard box for your eyes and hands, and turn on the UV lamp being careful not to damage your eyes. If the lichen cortex has xanthones it will fluoresce as a dull to bright orange or yellow color. If the lichen contains alectoronic acid or other subtler medullary chemicals, you will need to first flake off some of the cortex to expose the medulla before conducting the UV test. Alectoronic acid and other medullary chemicals turn a subtle to bright white or "ice blue" under UV, depending on the concentration. This can be a confusing spot test if the results are not obvious, just know that a dull or vibrant purple color indicates a negative UV reaction.

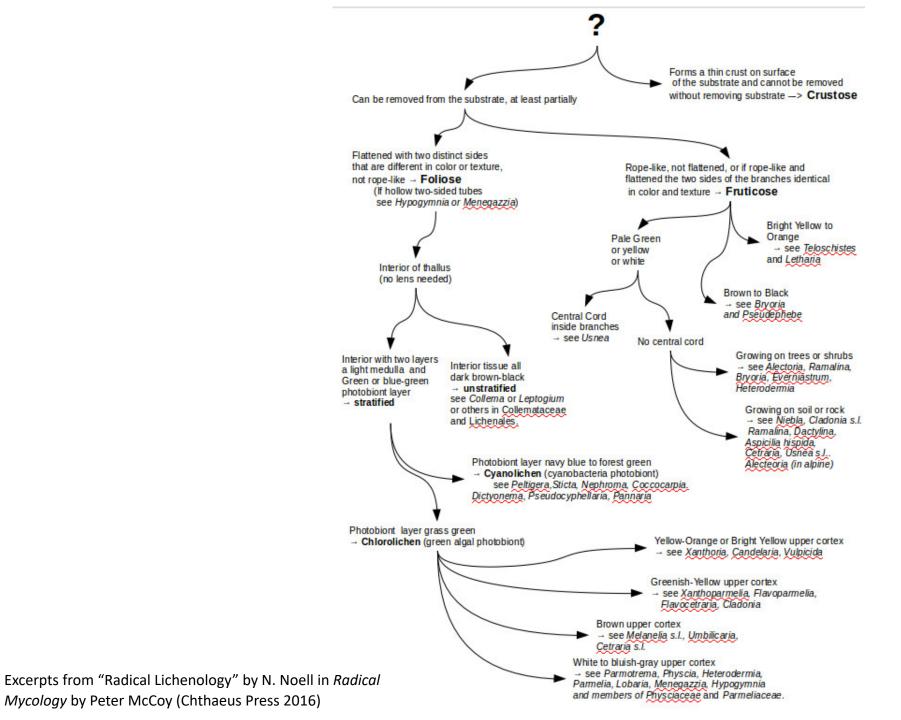






Spot test gear

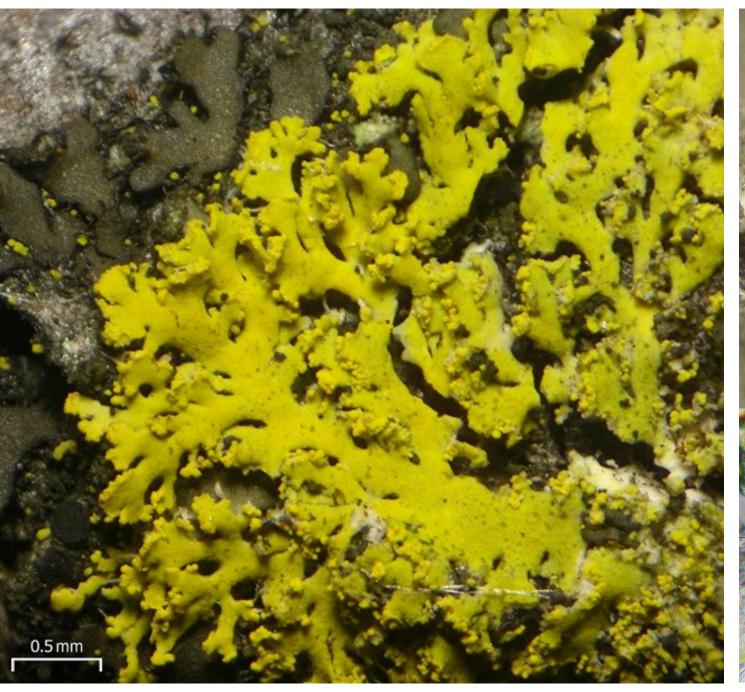
My preferred capillary tubes are made by Fisher Scientific (70ul, product number 22-260-943). You can get 100 for ten bucks. Before you use one, first point by holding the middle o the glass capillary tube over small flame until the glass i soft. Then pull from oppositi ends to break the tube at the center. Using sand paper or a rough surface, gently rub the ale. The capillary tube will pull chemicals up inside using capillary force and will pour them onto the lichen when the tip touches the thallus.









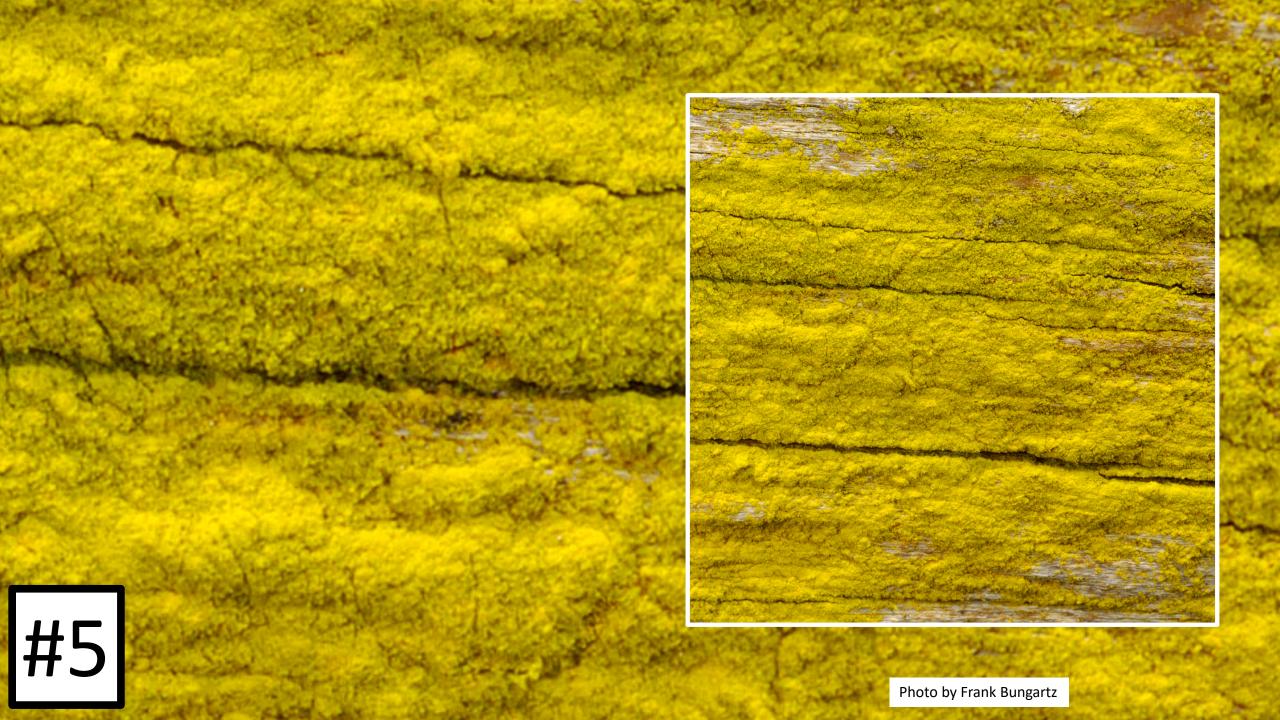


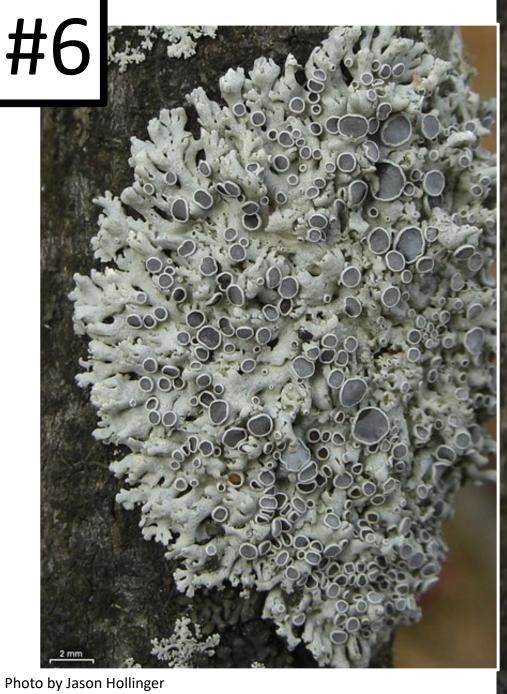


25μm

Photos by Jason Hollinger

#4





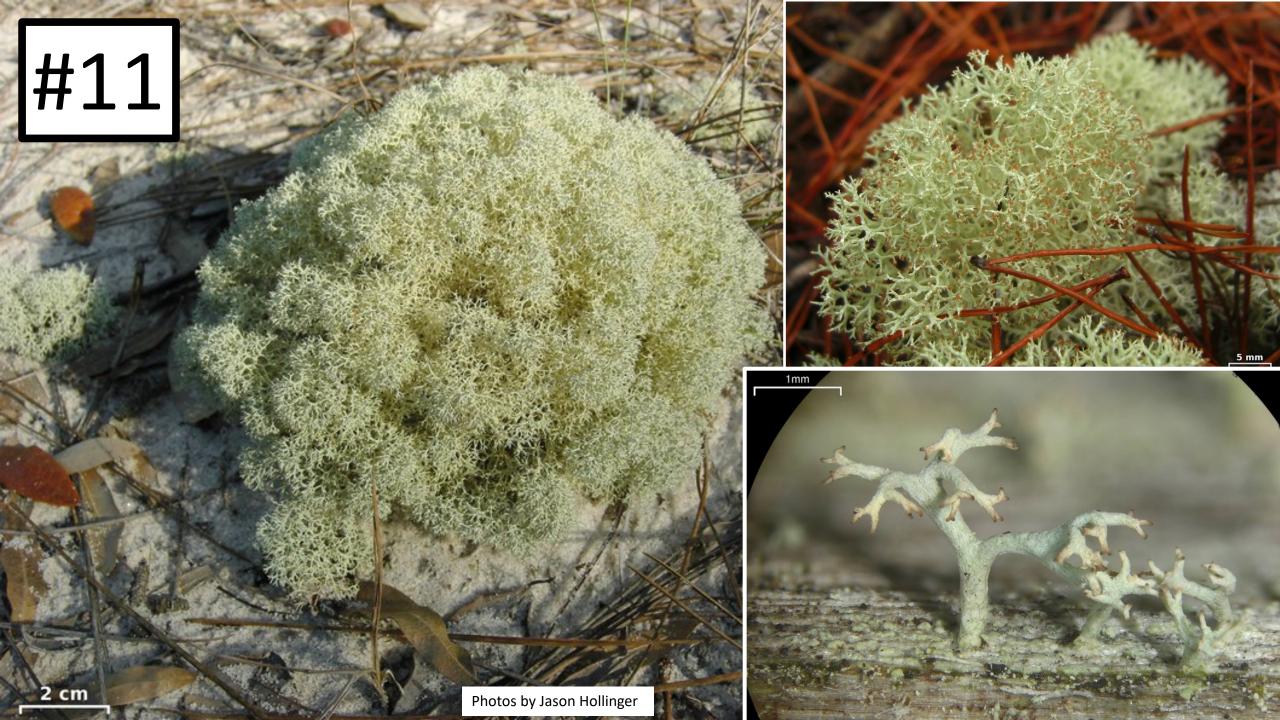




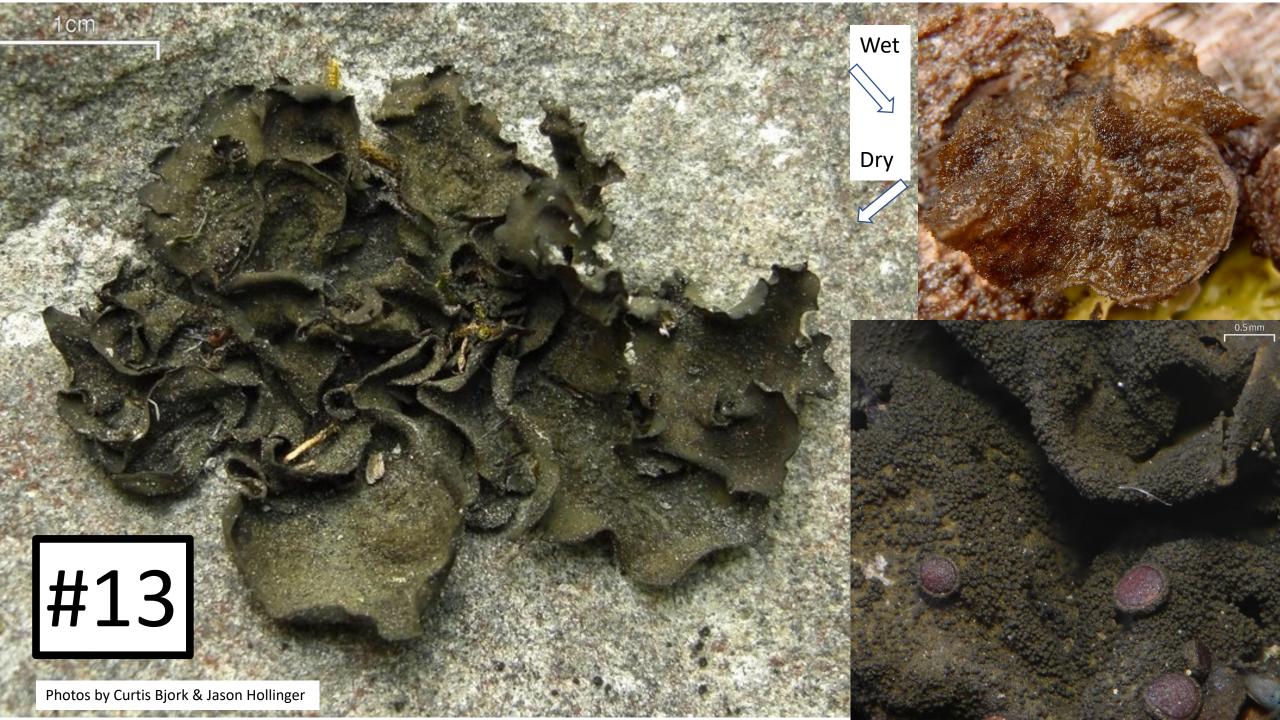


















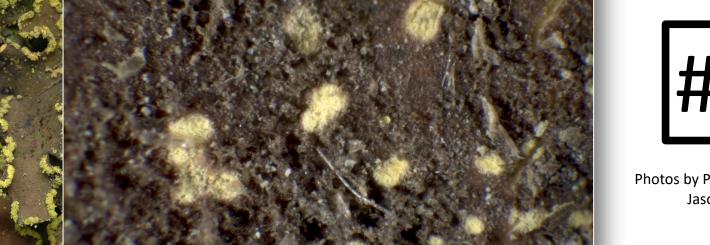






Photos by Jason Hollinger

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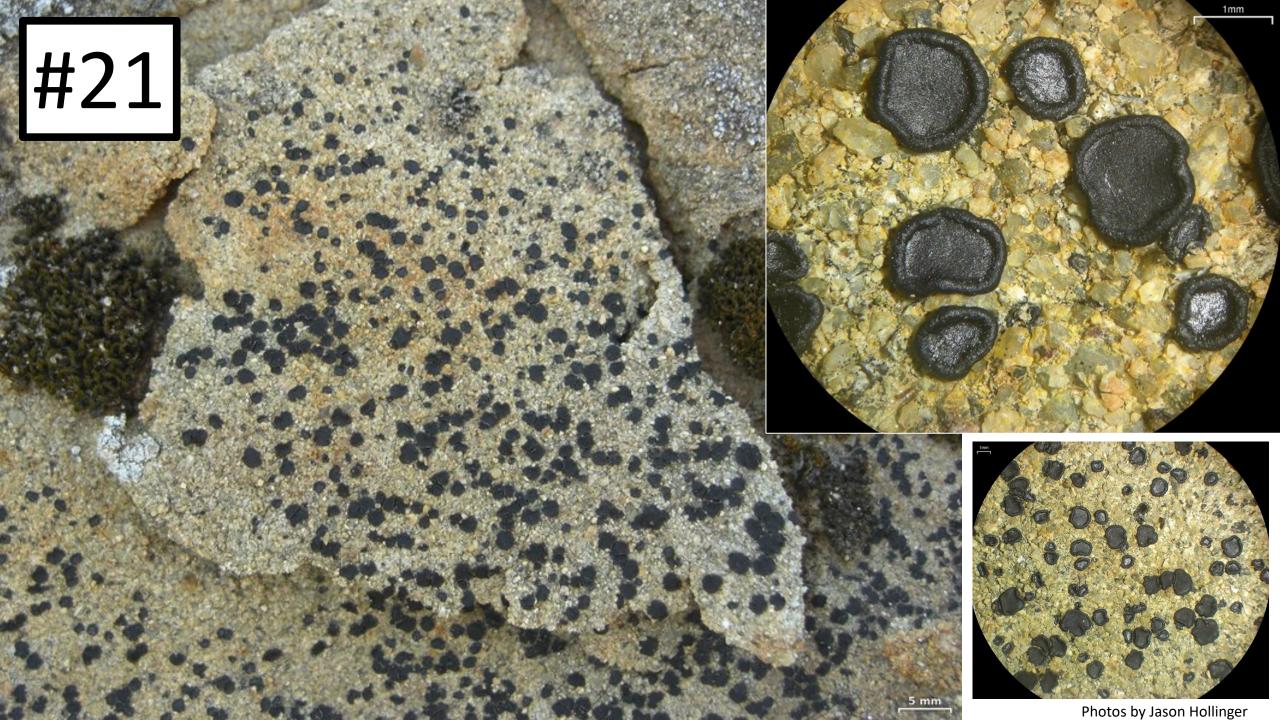


#19

Photos by P.L. Nimis, Stridvall & Jason Hollinger



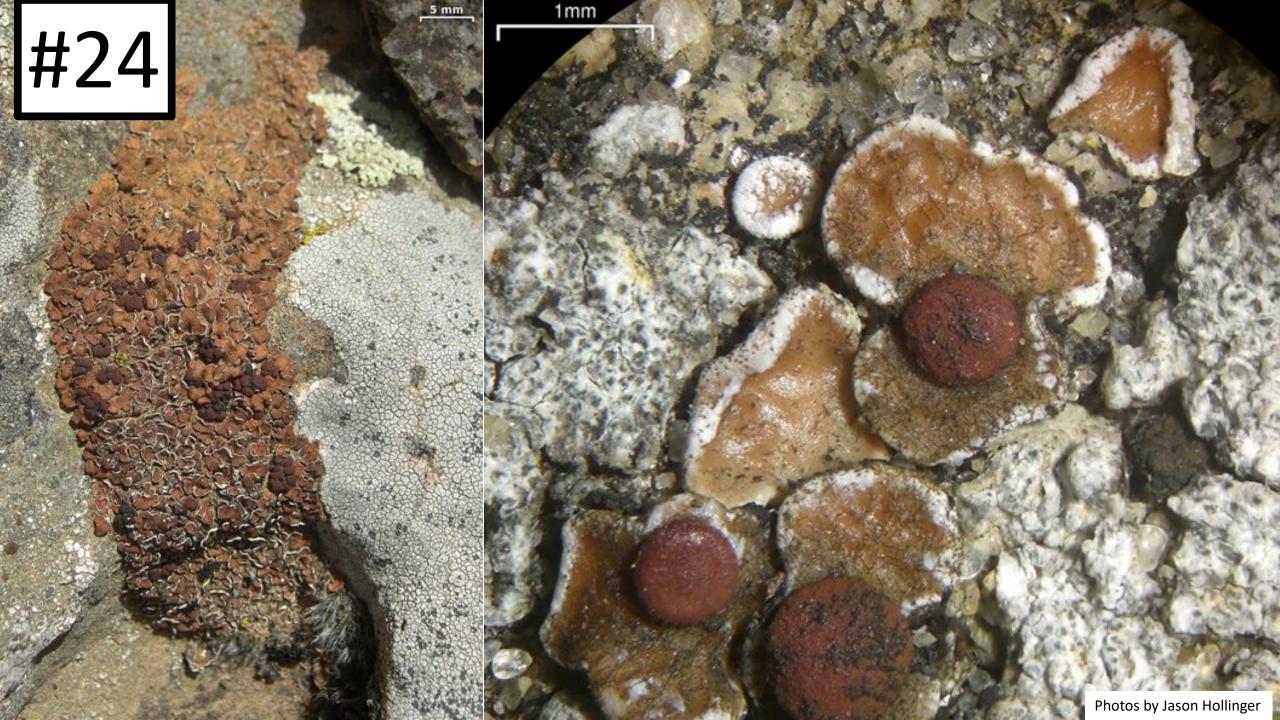








Photos by Jason Hollinger



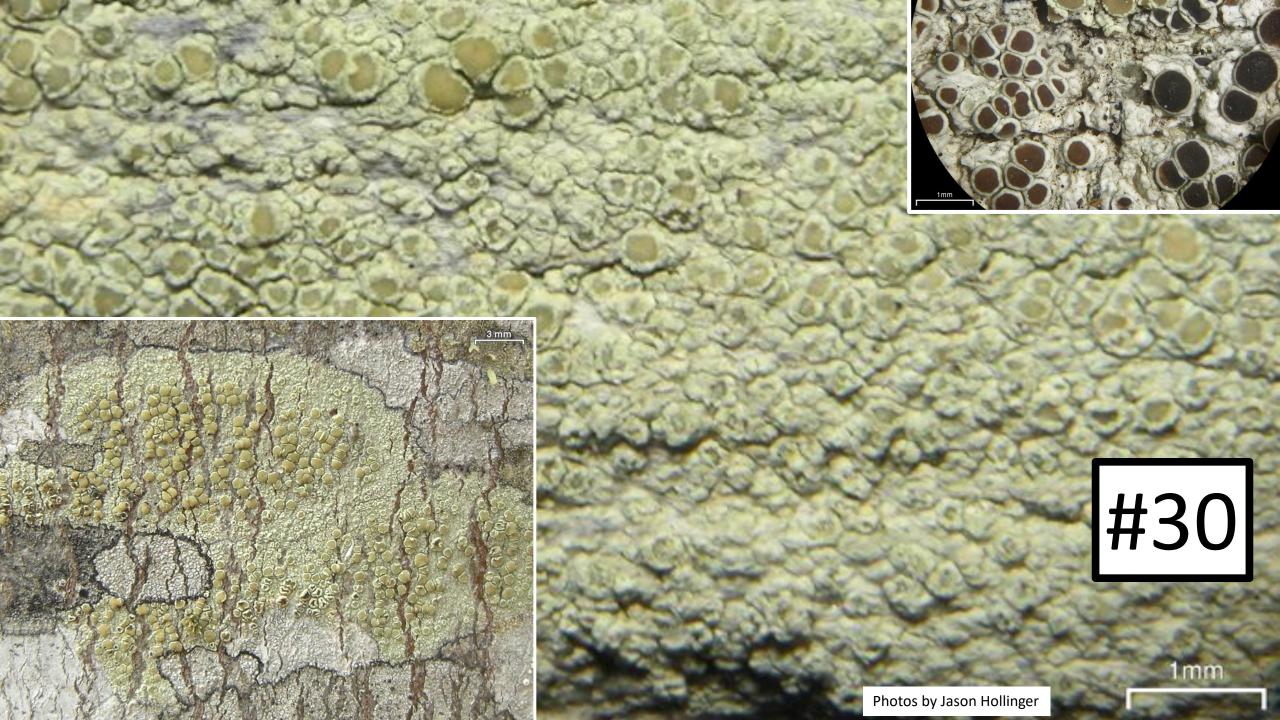


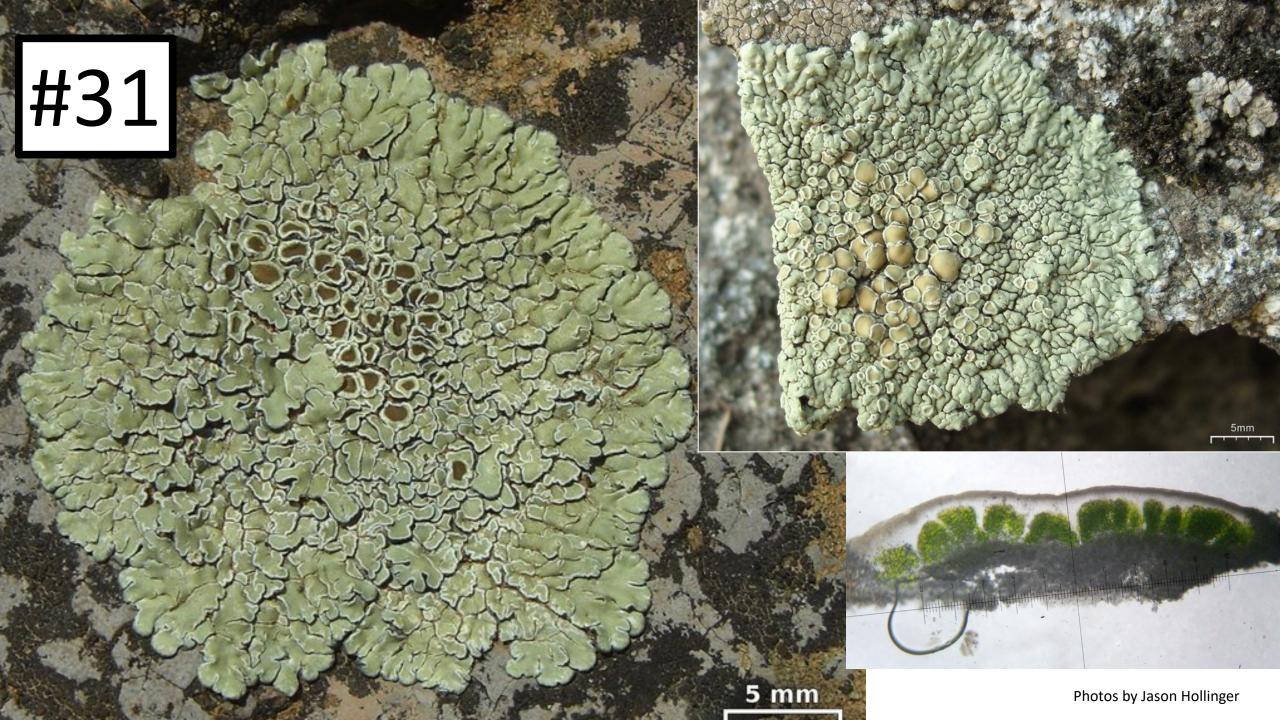




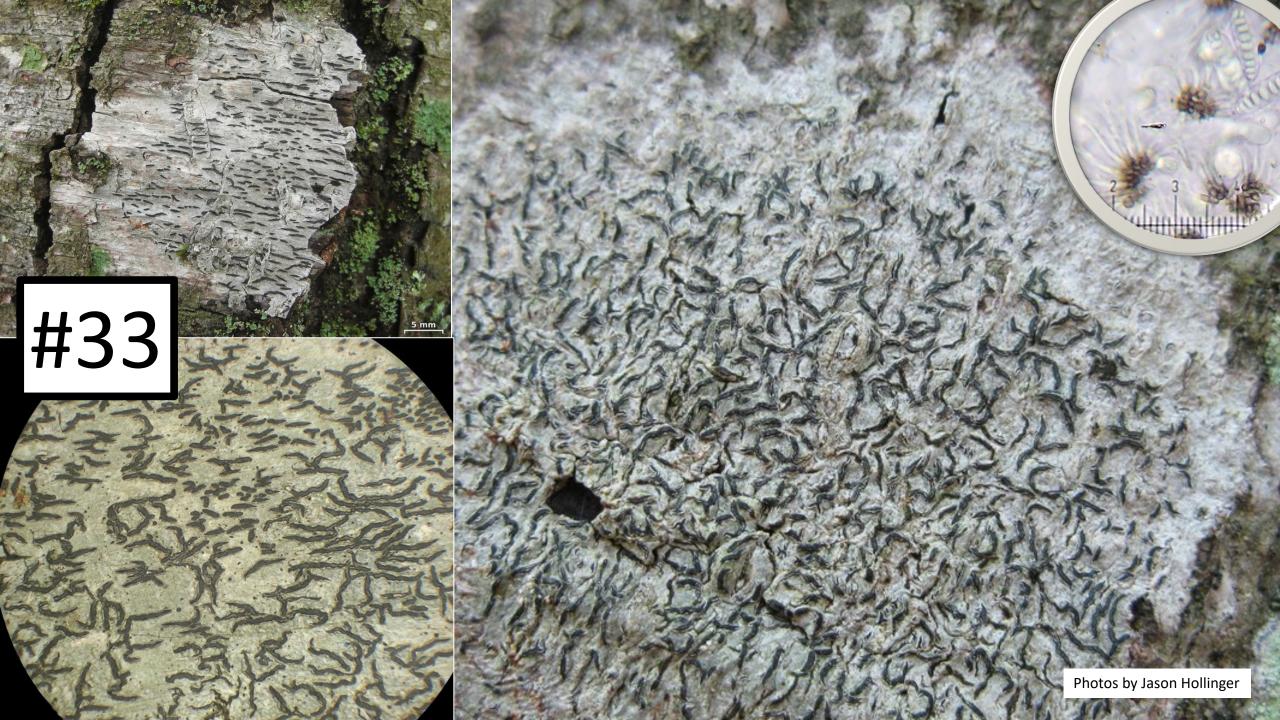






















KEY TO SECTIONS

La. Thallus fruticose, in form resembling an erect to pendent shrub, rope, stalk, or strap, with round to flattened bra upper cortex	nches, generally lacking a distinct lower cortex that is differentiated from the
1b. Thallus crustose, foliose, or squamulose, usually flattened, either closely attached to the substrate or with a disti	inct lower cortex, or flattened, lobe-like, and ± adnate to the substrate 2
2a. Thallus foliose to squamulose, of ± flattened, typically horizontally spreading, distinct lobes, often with a dististructure; rhizines or tomentum often present	inct lower cortex; upper & lower surfaces usually different in color and/or
3a. Thallus foliose, typically of branched, radiating lobes or rosettes	
2b. Thallus crustose, usually closely adherent to the substrate and often nearly inseparable from it; lower cortex l	acking; rhizines and tomentum absent 4
4a. Thallus routinely sterile, even in well developed specimens KEY D sterile crusts 4b. Thallus fertile, producing ascomata with asci and, usually, ascospores	
5a. Ascomata on distinct stalks, or slender, stalk-like structures (hyphophores) present	
7a. Apothecia elongate or branched, lirelliform to stellate, typically at least twice as long as wide 7b. Apothecia ± circular, sometimes slightly irregular or substellate in outline, but prevailingly isodia	

Keys by Douglass Ladd in **OZARK LICHENS**: Enumerating the lichens of the Ozark Highlands of Arkansas, Illinois, Kansas, Missouri, and Oklahoma; April 2018 Draft [revised and expanded from October 2005 draft]

KEY A: FRUTICOSE LICHENS (including submacroscopic filamentous taxa)

1a. Thallus flattened to terete, not filamentous, the main branches > 0.3 mm broad2
1b. Thallus filamentous, the main branches < 0.2 mm broad
2a. Thallus greenish gray to blue-gray, green, or grayish orange, not gelatinous; stratified with a distinct algal layer; largest branches >0.5 mm wide; photobiont <i>Trebouxia</i> 3
3a. Thallus branches flattened, not terete4
4a. Thallus grayish to orange, at least locally K+ magenta (parietin); usnic acid absent <i>Teloschistes chrysophthalmus</i>
4b Thallus greenish, K-, usnic acid present
3b. Thallus branches thicker, subterete to terete
5a. Thallus branches solid, not hollow6
6a. Thallus branches angular; medulla loose and cottony; fibrils absent Evernia mesomorpha
6b. Thallus branches terete, with a distinct central cord; fibrils present Usnea
5b. Thallus branches hollow
7a. Squamules often present; persistent granular primary thallus lacking; podetia and substrate various
7b. Squamules absent; persistent granular primary thallus evident; podetia gray, to 7 mm tall, dark-tipped; on exposed sandstone <i>Pycnothelia papillaria</i>
2b. Thallus black to dark olive brown, becoming gelatinous when wet, or threadlike and 0.2 mm wide; not stratified with a distinct algal layer; photobiont cyanobacteria or <i>Trentepohlia</i> .8
8a. Isidiate; main thallus branches >2 mm wide, black, sometimes pruinose, flattened, umbilicate to straplike9
8a. Isidiate; main thallus branches >2 mm wide, black, sometimes pruinose, flattened, umbilicate to straplike9 9a. Thallus black, not pruinose, typically subumbilicate; wet thallus <250 µm thick tichinella
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KEY B: FOLIOSE LICHENS

la. Thallus gelatinous when wet, black to dark slate gray or brown, lacking a distinct algal layer, upper and lower surfaces similar; photobiont cyanobacterial
2a. Thallus distinctly foliose, attached to the substrate at multiple locations, ± horizontally spreading and appressed to substrate
3a. Lobes extremely narrow, < 0.2 mm broad
3b. Lobes > 0.2 mm broad4
4a. Thallus dull above, black to brownish or olive; upper surface composed of loosely aggregated hyphae5
5a. Thallus distinctly foliose; ascospores 1+ septate; asci with IKI+ blue apical dome Collema
5b. Thallus subcrustose, thin and membranaceous; ascospores simple; asci IKILempholemma polyanthes
4b. Thallus sublustrous above, slate gray to rich brown, upper surface composed of a layer of ± isodiametric cellscells
2a. Thallus subfruticose to squamulose, typically attached to the substrate at a single point, ascending to umbilicate6
6a. Thallus of small subterete branches < 0.6 mm wideSynalissa symphorea
6b. Thallus of flattened straplike to umbilicate lobes mostly > 1 mm wide7
7a. Thallus of grayish pruinose ± straplike lobes
7b. Thallus epruinose, umbilicate to broadly squamulose Lichinella nigritella
Lb. Thallus not gelatinous, variously colored, with a distinct algal layer, upper and lower surfaces usually different colors; photobiont various8
8a. Upper cortex tinted with orange or yellow, ranging from yellow green or sea green to lemon yellow or orange9
9a. Upper cortex golden yellow to orange, K+ magenta (parietin)10
10a. Thallus esorediate, closely adnate; rhizines lacking (simple hapters sometimes present)11
11a. Thallus bright orange, distinctly foliose; pruina, if present, inconspicuous; lower cortex present Xanthoria elegans
11b. Thallus bright to pinkish or yellowish orange; often whitish pruinose; lower cortex absent
10b. Thallus sorediate, appressed to suberect; rhizines presentXanthomendoza
9b. Upper cortex lemon yellow to yellowish green or sea green, K12
12a. Upper cortex lemon yellow (sometimes greenish yellow in extremely shaded populations); lower cortex pale, rhizinate; thallus lobes < 0.4 mm wide Candelaria
12b. Upper cortex yellowish green; lower cortex various, but if thallus lobes < 0.5 mm wide then thallus subcrustose and lower cortex lacking rhizines13
13a. Thallus subcrustose to squamulose-umbilicate, either of discrete, bullate, subumbilicate areoles < 5 mm diameter or with areolate central portions and closely appressed marginal lobes; rhizines lacking14
14a. Thallus of suberect, swollen, bullate, subumbilicate areoles; lower surface pale brown, corticate
14a. Thailus of suberect, swoller, builate, suburnomicate areoles, lower surface pale brown, corticate
15a. On siliceous rock; apothecia black; ascospores brownish, 2-celled
15a. On sinceous rock, apothecia black, ascospores brownish, 2-cened
130. Substrate various, typically calcherous, apothecia tall to brown, ascospores hyaline, simple Lecunoru

13b. Thallus foliose, with a distinct rhizinate lower cortex; thallus not areolate16	В
16a. Thallus loosely adnate and convoluted; medulla bright yellow; lower cortex yellowish Vulpicida viridis	
16b. Thallus closely adnate, ± flat; medulla white; lower cortex whitish to tan, brown, or black17	
17a. Lobes broad, apically rounded, some > 3.5 mm wide; upper cortex dull to lustrous18	
18a. Isidiate or sorediate; apothecia rare19	
19a. Isidiate, the isidia sometimes breaking to appear apically sorediate20	
20a. Isidia fine, cylindrical; thallus margins ciliate	
20b. Isidia coarse, pustular, appearing apically sorediate with age; marginal cilia absentFlavoparmelia baltimorensis	
19b. Sorediate, the soredia sometime originating from pustules, but coalescing to form laminal sorediaFlavoparmelia caperata	
18b. Diaspores absent; apothecia commonFlavoparmelia rutidota	
17b. Lobes narrower and typically more elongate, prevailingly < 3.5 mm wide; upper cortex ± lustrousXanthoparmelia	
8b. Upper cortex brown, gray, or bluish or greenish gray, without yellowish tints (some species may have yellow soredia or medullary tissue)	21
21a. Upper cortex brown to brownish gray, K22	
22a. Lower surface tomentose, often felt-like23	
23a. Lower surface with distinct raised or darkened veins or regularly maculate with large white rounded patches	
23b. Lower surface lacking raised or darkened veins or maculations24	
24a. Lower surface with abundant small pores25	
25a. Thallus with marginal yellow soredia (often also marginally squamulose); lower surface with shallow, irregular punctations (pseudocyphellae)P.	seudocyphellaria aurat
25b. Thallus with abundant fimbriate-dissected marginal lobules; lower surface uniformly tan, with deep, circular pores (cyphellae)Sticta carolir	niensis
24b. Lower surface lacking pores; soredia, if present, not yellow26	
26a. Thallus lustrous, deep brown, with laminal, flattened, isidia-like lobules; apothecia on underside of lobe tips Nephroma helveticum	
26b. Thallus dull, gray brown, lobules, if present, marginal and not strongly flattened; apothecia on upper surface	
27a. Apothecia without a thalline marginSantessoniella crossophylla 27. Apothecia with a distinct thalline margin28	
28a. Thallus squamulose to subfoliose, grayish, with abundant whitish marginal zones; hymenium ultimately IKI+ reddish brown (sometime initially bi	lue green); ascospore
smooth sheaths Fuscopannaria	
28b. Thallus foliose, brown, essentially without whitish marginal zones; hymenium persistently IKI+ blue (only around asci); ascospores with ± short a	piculus and rough
sheaths Pannaria	
22b. Lower surface bare or rhizinate, lacking tomentum	
29a. Thallus umbilicate and centrally attached or of aggregated, subumbilicate thalli; well-defined branching lobes absent or abortive; diaspores absent	30
30a. Thallus typically > 5 cm broad; lower surface rhizinate; perithecia absent Umbilicaria mammulata	
30b. Thallus rarely > 5 cm broad; lower surface various; perithecia present and typically abundant Dermatocarpon	
29b. Thallus characteristically foliose, with branched lobes; diaspores present or absent; apotheciate or sterile31	
31a. Sorediate32	
32a. Upper cortex densely pruinose, at least near lobe tips	
32b. Upper cortex not pruinose33	
33a. Lower cortex black	
33b. Lower cortex pale34	